

HETEROSIS AND COMBINING ABILITY IN LATE MATURING QPM HYBRIDS IN MAIZE (*ZEA MAYS* L.)

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Abstract: Fifteen maize inbred were crossed as lines to three testers(HKI-193-1, CLQRCYQ-40-3-1 and HO6R-6136-68-1-1-6) in Line X Tester mating design to generate 45 F₁ crosses. The sixty seven genotypes including 45 F₁ hybrids along with their 18 parents and 4 checks were evaluated for five qualitative traits namely starch content, oil content, protein content, tryptophan content and lysine content in Randomized Block Design to estimate the General Combining Ability (GCA), Specific Combining Ability (SCA) and Heterosis of F₁ crosses. In the Line x Tester analysis, significant variability was observed among the genotypes for five traits. The hybrids L₃ x T₃ and L₅ x T₃ exhibited maximum mean values for oil content and hybrid L₁₀ x T₁ exhibited maximum mean values for starch content. The maximum mean value for protein content was depicted by hybrid L₆ x T₂, whereas hybrid L₁₀ x T₃ exhibited maximum mean value for tryptophan content. The hybrid L₇ x T₃ exhibited maximum mean values for lysine content.

Keywords: Combining ability, Grain yield, Heterosis, Quality protein maize

INTRODUCTION

Maize (*Zea mays* L.; 2n=20) is the third most important cereal crop among the cereals grown in India and is one of the promising crops for food, feed, fodder and industrial utilization. However, its protein is deficit in essential amino acids particularly, lysine and tryptophan. To overcome this deficiency, Quality Protein Maize (QPM) donors with sufficiently higher quantity of lysine and tryptophan have been developed at CIMMYT Mexico (Vasal, 1999). Maize protein is characterized by high level of glutamic acid and leucine. Lysine (1.2 per cent of protein) and tryptophan (0.4 per cent) are limiting amino acids in maize. This is due to the fact that major storage protein is a prolamine fraction zein, which forms up to 50-60 per cent of the storage protein. Zein consists of a group of hydrophobic proteins, completely devoid of lysine and tryptophan (Inglett, 1970). Hence genetic manipulation for improved nutritional value, particularly, protein quality was considered as a noble goal. This effort was stimulated by the 1963 discovery that a little known mutant maize contained proteins that are nearly twice as nutritious as those found in normal maize called "opaque 2 maize", its protein had a nutritive value about 90 per cent of that of proteins found in skim milk. The discovery of this mutant and subsequently its modifier were considered remarkable and led to the concept of "Breeding for Quality Protein Maize (QPM)" (Vasal, 1999). The lysine levels in normal and QPM maize average 2.0 per cent and 4.0 percent of total protein respectively, but range across genetic backgrounds from 1.6 - 2.6 per cent in normal maize and 2.7 - 4.5 percent in their o2 converted counterparts (Moro *et al.*, 1996). The lysine content of QPM in whole grain is about

0.33 per cent to 0.54 per cent, with the average 0.38 per cent, and is 46 per cent higher than normal maize. The tryptophan content is 0.08 percent, 66 per cent higher than normal maize. (Ortega *et al.*, 1986; Sproule *et al.*, 1988; Osei *et al.*, 1999). Normal maize contributes up to a third or more of the crude protein content of chicken diets. On the other hand, maize is low in protein in addition to its general deficiency in essential amino acids, particularly lysine and tryptophan. An understanding of the genetic architecture of parents and their mode of inheritance will greatly assist the breeder to formulate appropriate breeding methodologies to incorporate the traits in question into an otherwise desirable variety. Various biometrical approaches are available to access the breeding value of the traits under transfer. Line X tester analysis is one of those employed by which the genetic architecture of given character, the combining ability and heterosis could be understood. Most efficient use of such materials would be possible only when adequate information on the amount and type of genetic variation, combining ability effects and heterotic effects in the materials is available. In this context, L x T analysis (Kempthorne, 1957) has been widely used for evaluation of inbred lines by crossing them with testers. The present investigation was undertaken for estimation of combining ability and heterosis of normal inbred lines with QPM donors as a tester for initiating a successful quality protein maize conversion programme.

MATERIAL AND METHOD

The present investigation was carried out during the Kharif season of 2013-2014 at Instructional farm, Rajasthan College of Agriculture, Maharana

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Pratap University of Agriculture and Technology, Udaipur, India. The parental material for present investigation comprised of 15 inbred lines, 3 testers and 4 checks. These were selected on the basis of wide diversity for different metric traits. The inbred lines were developed and maintained at the Centre of AICRP on Maize, Udaipur, whereas, testers for investigation viz., HKI-193 was collected from CCSHAU, Uchani, Karnal, CLQRCYQ-40-3-1 was collected from DMR, New Delhi and EIQ-102 from AICRP on Maize, Udaipur. The experimental material consisted of 45 crosses made during Rabi 2013, involving 15 lines, 3 testers along with 4 checks sown in a Randomized Block Design with three replications in Kharif 2013-2014. Each genotype was planted in a single row plot of 4 m length having a uniform inter and intra row spacing of 60 cms and 20 cms respectively. Two seeds per hill was planted and later on one plant was thinned (if necessary) from each hill to maintain the optimum plant population. Border rows were planted to avoid border effect. The data was subjected to the estimation of heterosis over better parent and standard variety (Fonseca and Patterson, 1968).

RESULT AND DISCUSSION

The analysis of variance revealed that mean squares due to parents and crosses were highly significant for all the characters (Table 1). This reflected presence of adequate genetic variability in the experimental material included under study. Similarly, significant mean squares due to parents vs. crosses indicated presence of average heterosis for all the characters. Considerable amount of heterosis was observed for all the characters under study, however the magnitude varied with characters (Table 2).

In the case of oil content, ten hybrids exhibited positive significant economic heterosis over the best check Pratap QPM hybrid-1. The maximum positive significant economic heterosis was depicted by the hybrid $L_5 \times T_3$ (20.38 per cent) followed by hybrid $L_3 \times T_3$ (20.32 per cent) and hybrid $L_1 \times T_3$ (19.67 per cent). Poneleit and Bauman (1970), Oropeza *et al.* (1984), Joshi *et al.* (1985), Lico (1989), Ivanov (1990) reported economic heterosis for oil content. For starch content, only two hybrids viz., $L_9 \times T_1$ (2.67 per cent) and $L_4 \times T_2$ (2.56 per cent) exhibited positive significant economic heterosis over the best check PMH-4 (Table 2). Dubey *et al.* (2009), Premalatha and Kalamani (2010), Pavan *et al.* (2011), Abuali *et al.* (2012), Singh *et al.* (2013) reported economic heterosis for seed oil content and starch content in maize.

The significant economic heterosis in positive direction was observed in ten hybrids for protein content with the magnitude ranged from 2.21 ($L_{13} \times T_1$) to 14.47 per cent ($L_6 \times T_2$). The maximum positive significant economic heterosis was expressed by hybrid $L_6 \times T_2$ (14.47 per cent) followed by hybrid $L_5 \times T_1$ (13.97 per cent) and hybrid $L_6 \times T_3$ and $L_9 \times T_3$ (6.37 per cent). The best check for this trait was PMH-4. In the case of tryptophan content, three hybrids viz., $L_{10} \times T_3$ (10.59 per cent), $L_{13} \times T_1$ (9.75 per cent) and $L_5 \times T_1$ (3.39 per cent) exhibited positive significant economic heterosis over the best check Pratap QPM hybrid-1. For lysine content, only one hybrid $L_7 \times T_3$ (11.20 per cent) exhibited positive significant economic heterosis over the best check HQPM-7 (Table 3). The estimates of gca effects revealed that inbred line L_6 has been found good general combiner for majority of traits viz., protein content, tryptophan content, lysine content and oil content.

Table 1. Analysis of variance for combining ability for fourteen traits in maize

Source of Variation	d.f.	Oil content	Starch content	Protein content	Tryptophan content	Lysine content
Replication	2	0.0017915	1.11136	0.019464	0.00086**	0.00138955
Genotype	66	1.15901**	11.2226**	2.85890**	0.02228**	0.148883**
Parents	17	0.38630**	11.4575**	0.655283**	0.01862**	0.114285**
Parent v/s Crosses	1	0.23489**	26.1513**	63.1113**	0.00583**	0.012038**
Crosses	44	1.5540**	11.3381**	2.51028**	0.02581**	0.174887**
Lines	14	1.01664**	6.47167**	3.16054**	0.02883**	0.126802**
Tester	2	4.51353**	28.9602**	0.22196**	0.05651**	0.059644**
Line x Tester	28	1.61137**	12.5127**	2.34860**	0.02211**	0.207161**
Error	88	0.0009633	0.742169	0.016409	0.0001430	0.0005053
Variance						
σ^2_L		-0.066082	0.67123	0.090216	0.0007461	-0.0089287
σ^2_t		0.064492	0.3655	-0.047259	0.0007644	-0.0032782
σ^2_{GCA}		-0.002918	-0.059814	0.0082337	0.0001884	-0.0016436
σ^2_{SCA}		0.5368	3.9235	0.7774	0.0073242	0.068885
$\sigma^2_{SCA/GCA}$		183.9616	65.595011	94.4168	38.875796	41.911049

*, ** Significant at 5 % and 1 % level of significance respectively.

Table 2. Extent of heterosis for oil content (%) and starch content (%)

SN.	Crosses	Oil content (%)			Starch content (%)		
		Het	Hb	EH	Het	Hb	EH
1.	L1 x T1	10.19**	3.27**	1.88**	1.58	1.09	-
2.	L2 x T1	-3.56**	-	-	5.03**	2.12	-
3.	L3 x T1	-1.74**	-	-	3.52**	3.18**	0.70
4.	L4 x T1	15.22**	14.16**	-	1.57	0.32	-
5.	L5 x T1	2.75**	0.34	-	2.24*	2.16	-
6.	L6 x T1	8.99**	2.46**	0.41	-1.68	-	-
7.	L7 x T1	-13.47**	-	-	0.94	0.69	-
8.	L8 x T1	-8.64**	-	-	2.63**	2.55*	-
9.	L9 x T1	8.33**	3.81**	-	2.28*	2.12	-
10.	L10 x T1	13.87**	13.75**	-	4.69**	3.51**	2.67**
11.	L11 x T1	7.58**	1.51**	-	-1.51	-	-
12.	L12 x T1	-9.52**	-	-	1.61	1.34	-
13.	L13 x T1	10.43**	4.43**	1.06*	2.55*	-	-
14.	L14 x T1	13.42**	13.34**	-	-1.21	-	-
15.	L15 x T1	-3.29**	-	-	0.80	0.12	-
16.	L1 x T2	-22.98**	-	-	1.32	-	-
17.	L2 x T2	12.76**	4.68**	1.17**	2.50*	-	-
18.	L3 x T2	20.03**	18.13**	1.00*	2.63**	1.40	1.40
19.	L4 x T2	-5.75**	-	-	5.42**	2.56**	2.56**
20.	L5 x T2	3.84**	3.48**	-	-1.25	-	-
21.	L6 x T2	1.33**	-	-	1.14	-	-
22.	L7 x T2	-15.19**	-	-	-2.26*	-	-
23.	L8 x T2	7.29**	-	-	-0.07	-	-
24.	L9 x T2	11.32**	4.62**	-	-0.84	-	-
25.	L10 x T2	-3.34**	-	-	-4.87**	-	-
26.	L11 x T2	-23.87**	-	-	-0.98	-	-
27.	L12 x T2	31.54**	28.34**	11.69**	0.24	-	-
28.	L13 x T2	-3.27**	-	-	3.19**	-	-
29.	L14 x T2	-14.77**	-	-	-6.13**	-	-
30.	L15 x T2	-7.00**	-	-	0.62	-	-
31.	L1 x T3	29.81**	21.31**	19.67**	-5.37**	-	-
32.	L2 x T3	9.27**	3.10**	-	2.09*	1.09	-
33.	L3 x T3	40.53**	40.34**	20.32**	-2.19*	-	-
34.	L4 x T3	20.40**	19.66**	2.58**	1.06	0.44	-
35.	L5 x T3	43.36**	40.41**	20.38**	6.09**	4.07**	1.06
36.	L6 x T3	7.13**	0.42	-	2.83**	0.59	-
37.	L7 x T3	0.42	-	-	2.75**	1.11	-
38.	L8 x T3	-8.00**	-	-	1.27	-	-
39.	L9 x T3	1.21**	-	-	-2.20*	-	-
40.	L10 x T3	22.42**	22.17**	5.17**	-8.01**	-	-
41.	L11 x T3	6.93**	0.60	-	7.35**	6.09**	1.50
42.	L12 x T3	-11.49**	-	-	3.65**	2.03	-
43.	L13 x T3	6.89**	0.79	-	10.29**	6.38**	-
44.	L14 x T3	1.40**	1.16*	-	-2.07*	-	-
45.	L15 x T3	-13.48**	-	-	2.14*	0.96	-

*, ** Significant at 5 % and 1 % level of significance respectively.

Table 3. Extent of heterosis for protein content (%), tryptophan content (%) and lysine content (%)

SN.	Crosses	Protein %			Tryptophan %			Lysine %		
		Het	Hb	EH	Het	Hb	EH	Het	Hb	EH
1.	L1 x T1	26.20**	20.52**	-	-7.20**	-	-	-8.04**	-	-
2.	L2 x T1	0.29	-	-	8.83**	0.00	-	3.66**	-	-
3.	L3 x T1	28.53**	25.20**	2.98**	9.55**	-	-	4.85**	-	-
4.	L4 x T1	23.43**	20.77**	-	19.52**	15.03**	-	17.93**	16.64**	-
5.	L5 x T1	37.13**	35.74**	13.97**	26.10**	7.49**	3.39**	5.96**	-	-
6.	L6 x T1	10.16**	6.46**	-	0.27	-	-	-4.24**	-	-
7.	L7 x T1	25.21**	24.86**	3.28**	16.57**	10.67**	-	-5.54**	-	-
8.	L8 x T1	9.14**	6.78**	-	34.46**	24.38**	-	-7.82**	-	-
9.	L9 x T1	7.49**	3.99**	-	19.03**	15.20**	-	16.98**	15.14**	-
10.	L10 x T1	10.95**	9.33**	-	21.14**	11.58**	-	15.43**	11.27**	-
11.	L11 x T1	5.00**	-	-	17.51**	7.22**	-	12.54**	7.44**	-
12.	L12 x T1	11.08**	9.41**	-	7.07**	-	-	3.51**	-	-
13.	L13 x T1	27.46**	24.27**	2.21*	30.81**	9.75**	9.75**	-9.76**	-	-

14.	L14 x T1	-7.28**	-	-	17.26**	4.39**	-	14.38**	8.44**	-
15.	L15 x T1	2.63*	0.43	-	9.83**	2.15	-	2.92**	-	-
16.	L1 x T2	48.37**	44.92**	13.76**	-11.23**	-	-	-10.06**	-	-
17.	L2 x T2	10.03**	3.99**	-	0.26	-	-	0.50	-	-
18.	L3 x T2	20.14**	19.75**	-	14.80**	14.80**	-	-5.37**	-	-
19.	L4 x T2	9.67**	9.53**	-	3.52*	-	-	4.06**	-	-
20.	L5 x T2	15.11**	11.37**	-	-22.93**	-	-	-5.58**	-	-
21.	L6 x T2	37.35**	29.81**	14.47**	7.02**	1.84	-	7.68**	2.46**	0.77
22.	L7 x T2	12.26**	9.39**	-	2.14	-	-	3.18**	-	-
23.	L8 x T2	27.39**	21.85**	4.76**	-10.24**	-	-	4.64**	1.73*	-
24.	L9 x T2	22.11**	20.86**	-	-10.08**	-	-	-9.55**	-	-
25.	L10 x T2	8.21**	4.23**	-	-3.63**	-	-	-4.07**	-	-
26.	L11 x T2	7.35**	-	-	20.00**	19.39**	-	8.24**	8.03**	-
27.	L12 x T2	9.86**	8.98**	-	-19.31**	-	-	-17.94**	-	-
28.	L13 x T2	18.20**	17.92**	-	-15.74**	-	-	-10.63**	-	-
29.	L14 x T2	30.60**	30.02**	2.98**	-31.67**	-	-	-0.55	-	-
30.	L15 x T2	4.73**	0.19	-	-10.47**	-	-	-13.07**	-	-
31.	L1 x T3	17.14**	13.02**	-	0.54	0.00	-	0.78	0.13	-
32.	L2 x T3	9.47**	4.71**	-	-12.77**	-	-	-12.27**	-	-
33.	L3 x T3	21.06**	19.18**	-	-13.39**	-	-	3.81**	0.62	-
34.	L4 x T3	5.85**	4.66**	-	3.35*	0.00	-	3.18**	0.13	-
35.	L5 x T3	15.13**	12.76**	-	-18.93**	-	-	-15.96**	-	-
36.	L6 x T3	34.92**	29.05**	13.80**	3.98**	-	-	-13.03**	-	-
37.	L7 x T3	20.64**	19.03**	-	28.37**	25.95**	-	35.79**	32.94**	11.20**
38.	L8 x T3	20.02**	16.21**	-	16.51**	1.08	-	14.02**	13.87**	-
39.	L9 x T3	44.60**	41.35**	13.80**	0.56	-	-	13.45**	9.58**	-
40.	L10 x T3	11.80**	9.01**	-	39.20**	37.37**	10.59**	5.28**	3.42**	-
41.	L11 x T3	10.78**	3.62**	-	17.68**	14.95**	-	-4.08**	-	-
42.	L12 x T3	-0.71	-	-	-5.85**	-	-	1.66**	-	-
43.	L13 x T3	18.13**	16.39**	-	7.36**	-	-	-7.56**	-	-
44.	L14 x T3	8.16**	7.28**	-	14.36**	8.78**	-	9.49**	5.75**	-
45.	L15 x T3	17.99**	14.26**	-	-9.43**	-	-	-1.15	-	-

*,** Significant at 5 % and 1 % level of significance respectively.

In the present study, parents were classified as high, average and low combiners based on their effects. Parents with desirable GCA effect (significantly different from zero) were considered as high combiners, while parents showing insignificant estimates were classified as average combiners. Low or poor combiners had significant but negative (undesirable) GCA effects. The estimates of gca effects revealed that inbred line L₆ has been found good general combiner for majority of traits viz., protein content, tryptophan content, lysine content and oil content. Ivy and hawlader (2000) and Hussain *et al.* (2003) also observed the similar phenomenon.

The SCA effects of the crosses for yield and different yield contributing characters are presented in Table 4. The significant positive sca effects for tryptophan content were observed in eighteen hybrids with the magnitude ranged from 0.02 (L₁₂ x T₁, L₁₅ x T₂ and L₈ x T₃) to 1.15 (L₅ x T₁). The maximum positive significant sca effects was recorded in hybrid L₅ x T₁

(1.15) followed by hybrid L₃ x T₂ (1.14) and hybrid L₁₀ x T₃ (1.13).

In the case of lysine content, the significant positive sca effects were observed in twenty-two hybrids with the magnitude ranged from 0.04 (L₁₅ x T₃) to 0.58 (L₇ x T₃). The maximum positive significant sca effects was recorded in hybrid L₇ x T₃ (0.58) followed by hybrid L₆ x T₂ (0.41) and hybrid L₅ x T₁ and L₈ x T₃ (0.23). Positive SCA indicate that lines are in opposite heterotic groups, while negative SCA effects indicate lines are in the same heterotic group.

In general, the GCA effects of the parents were not reflected in the SCA effects of the crosses in most of the studied traits. This is corroborated with the results of Debnath and Sarker (1987), Paul and Daura (1991), Debnath (1992) and Hussain *et al.* (2003). Besides, Deitos *et al.* (2000) reported that good general combining parent does not always show high sca effects in their hybrid combinations.

Table 4. GCA and SCA effects for Protein content (%), Tryptophan content (%), Lysine content (%), Oil content (%) and starch content (%)

SN	Genotype	Protein %	Tryptophan %	Lysine %	Oil content (%)	starch content %
1	T1	-0.08**	0.03**	0.02**	-0.01*	0.48**
2	T2	0.05*	-0.04**	-0.04**	-0.31**	0.44**
3	T3	0.03	0.01**	0.02**	0.32**	-0.93**
4	L1	0.63**	-0.07**	-0.22**	0.27**	-1.02**
5	L2	-0.52**	-0.04**	-0.11**	0.22**	-0.02

SN	Genotype	Protein %	Tryptophan %	Lysine %	Oil content (%)	starch content %
6	L3	0.26**	0.00	0.02*	0.54**	0.82**
7	L4	-0.51**	-0.01*	0.04**	0.05**	0.67*
8	L5	0.57**	-0.01**	0.01	0.29**	1.27**
9	L6	1.23**	0.04**	0.05**	0.25**	0.47
10	L7	0.24**	0.04**	0.13**	-0.55**	-0.11
11	L8	0.38**	-0.05**	0.01	-0.16**	0.51
12	L9	0.30**	-0.05**	-0.01	0.19**	-0.29
13	L10	-0.39**	0.08**	0.10**	0.15**	-1.23**
14	L11	-0.19**	0.09**	0.13**	-0.23**	0.33
15	L12	-0.95**	-0.04**	-0.04**	-0.20**	0.73*
16	L13	0.11*	0.09**	-0.17**	0.16**	-0.38
17	L14	-0.69**	-0.01**	0.20**	-0.38**	-1.84**
18	L15	-0.48**	-0.06**	-0.12**	-0.61**	0.07
19	L1 x T1	-0.13	-0.07**	-0.14**	0.28**	1.05
20	L2 x T1	-0.35**	-0.00	0.08**	-0.45**	0.69
21	L3 x T1	0.61**	-0.03**	0.02	-0.98**	0.94
22	L4 x T1	1.02**	-0.01	0.15**	0.30**	-1.13
23	L5 x T1	1.40**	0.15**	0.23**	-0.61**	-0.48
24	L6 x T1	-1.26**	-0.09**	-0.11**	0.22**	-1.96**
25	L7 x T1	0.67**	-0.06**	-0.50**	-0.17**	-0.12
26	L8 x T1	-0.62**	0.04**	-0.37**	-0.24**	0.43
27	L9 x T1	-1.17**	0.03**	0.16**	0.12**	1.16
28	L10 x T1	0.23**	-0.06**	0.17**	0.19**	4.27**
29	L11 x T1	-0.06	-0.08**	0.09**	0.60**	-2.35**
30	L12 x T1	0.53**	0.02*	0.13**	-0.58**	-0.56
31	L13 x T1	0.68**	0.09**	-0.09**	0.35**	-2.07**
32	L14 x T1	-1.23**	0.05**	0.09**	0.70**	0.80
33	L15 x T1	-0.31**	0.02	0.09**	0.28**	-0.66
34	L1 x T2	1.19**	0.03**	-0.01	-1.24**	1.93**
35	L2 x T2	0.14	0.08**	0.19**	0.59**	0.19
36	L3 x T2	-0.41**	0.14**	-0.06**	0.26**	1.43*
37	L4 x T2	-0.42**	0.04**	0.01	-0.51**	2.34**
38	L5 x T2	-0.77**	-0.06**	0.10**	-0.36**	-1.66**
39	L6 x T2	0.66**	0.09**	0.41**	0.02	0.85
40	L7 x T2	-0.74**	-0.01	-0.09**	-0.04	-1.12
41	L8 x T2	0.54**	-0.06**	0.14**	0.80**	-0.24
42	L9 x T2	-0.37**	-0.01	-0.30**	0.47**	0.21
43	L10 x T2	-0.33**	-0.07**	-0.14**	-0.45**	-0.82
44	L11 x T2	-0.18*	0.08**	0.19**	-0.81**	-1.01
45	L12 x T2	0.10	-0.02**	-0.27**	1.61**	-0.38
46	L13 x T2	-0.41**	-0.10**	0.07**	-0.16**	-0.62
47	L14 x T2	1.44**	-0.14**	-0.11**	-0.46**	-1.35*
48	L15 x T2	-0.46**	0.02*	-0.14**	0.30**	0.25
49	L1 x T3	-1.06**	0.04**	0.14**	0.96**	-2.98**
50	L2 x T3	0.22*	-0.07**	-0.28**	-0.13**	-0.87
51	L3 x T3	-0.20*	-0.11**	0.05**	0.72**	-2.37**
52	L4 x T3	-0.60**	-0.03**	-0.15**	0.20**	-1.20*
53	L5 x T3	-0.63**	-0.09**	-0.33**	0.97**	2.13**
54	L6 x T3	0.61**	-0.00	-0.30**	-0.23**	1.11
55	L7 x T3	0.07	0.08**	0.58**	0.21**	1.24*
56	L8 x T3	0.07	0.02*	0.23**	-0.55**	-0.18
57	L9 x T3	1.53**	-0.01	0.14**	-0.59**	-1.37*
58	L10 x T3	0.09	0.13**	-0.03*	0.26**	-3.45**
59	L11 x T3	0.24**	-0.01	-0.28**	0.22**	3.36**
60	L12 x T3	-0.62**	0.00	0.14**	-1.03**	0.94
61	L13 x T3	-0.28**	0.00	0.02	-0.19**	2.69**
62	L14 x T3	-0.20*	0.10**	0.02	-0.24**	0.54
63	L15 x T3	0.76**	-0.04**	0.04**	-0.58**	0.41
Standard error						
	Ti	0.02	0.00	0.00	0.01	0.15

SN	Genotype	Protein %	Tryptophan %	Lysine %	Oil content (%)	starch content %
	Lj	0.04	0.00	0.01	0.01	0.30
	Sij	0.09	0.01	0.02	0.02	0.59
	Ti-j	0.03	0.00	0.00	0.01	0.18
	Li-j	0.06	0.01	0.01	0.01	0.41
	Ti-Lj	0.05	0.00	0.01	0.01	0.31
	STi-Tj	0.11	0.01	0.02	0.03	0.73
	SiL-jL	0.12	0.01	0.02	0.03	0.81
	Sij-kl	0.12	0.01	0.02	0.03	0.83

*,** Significant at 5 % and 1 % level of significance respectively.

REFERENCES

- Abuali, A.I., Abdelmulla, A.A., Khalafalla, M.M., Idris, A.E. and Osman, A.M.** (2012). Combining ability and heterosis for yield and yield components in maize (*Zea mays* L.). *Australian Journal of Basic and Applied Sciences*. **6**(10): 36-41.
- Debnath, S.C. and K.R., Sarker** (1987). Genetic analysis of grain yield and some of its attributes in maize (*Zea mays* L.). *Thai J. Agric. Sci.*, **20**: 263-276.
- Debnath, S.C.** (1992). Analysis of heterosis in a 10 × 10 diallel cross of maize. *Bangladesh J. Agril. Sci.*, **19**: 161-164.
- Deitos, A., Arnhold, E., Mora, F. and Miranda, G.V.** (2006). Yield and combining ability of maize cultivars under different eco-geographic conditions. *Crop Breed. Appl. Boitec. Brazilian Society of Plant Breeding*, **6**: 222-227.
- Dubey, R.B., Joshi, V. N. and Verma, M.** (2009). Heterosis for nutritional quality and yield in conventional and non-conventional hybrids of maize (*Zea mays* L.). *Indian Journal of Genetics and Plant Breeding*. **69**(2): 109-114.
- Fonesca, S. and Patterson, F. L.** (1968). Hybrid vigour in a seven parent diallel cross in common wheat (*Triticum aestivum* L.). *Crop Science*. **8**: 85-88.
- Hussain, S.A., Amiruzzaman, M. and Hossain, Z.** (2003). Combining ability estimates in maize. *Bangladesh. J. Agril. Res.*, **28**: 435-440.
- Inglett, G.E.** (1970). Kernel structure, composition and quality In: Inglett GE (ed) corn: culture, processing, products. AVI, New York pp.468-502.
- Ivanov, S.** (1990). Effect of cultivation condition in maize in yield and control in protein and oil in the grain of F₁ hybrids. *Institute Pa Tsaravitsata, Kzezha. Bulgaria Bulletin Po Kukuruz*. pp. 311-325.
- Ivy, N.A. and Howlader, M.S.** (2000). Combining ability in maize. *Bangladesh J. Agril. Res.*, **25**: 385-392.
- Joshi, V.N., Sharma, G.S. and Ranwah, B.R.** (1985). Prospects of maize (*Zea mays* L.) as a new oil seed crop. In oil seed production, constraints and opportunities (Edited by H.C. Shrivastava and S. Bhaskaran). IBH Publi. Co., 529-533.
- Kempthorne, O.** (1957). An introduction to genetical statistics. John Wiley and Sons Inc., New York, pp. 323-331.
- Moro, G.L., Habben, J. E., Hamaker, B. R. and Larkins, B. A.** (1996). Characterization of the variability in lysine content for normal and opaque2 maize endosperm. *Crop Sci.* **36**: 1651-1659.
- Oropeza, E., Ortiz, L. and Bertorelli, D.E.** (1984). Assessment of nutritional quality of protein in six cultivars of maize (*Zea mays* L.). *Revista de la facultad Agronomic, Univ. Central de Venezuela*. **15**(3-5): 225-233.
- Ortega, E. I., Villegas, E. and Vasal, S. K.** (1986). A comparative study of protein changes in normal and quality protein maize during tortilla making. *Cereal Chem.*, **63**(5):446-451.
- Osei, S. A., Dei, H. K. and Tuah, A. K.** (1999). Evaluation of quality protein maize as a feed ingredient for layer pullet. *J. Anim. Feed Sci.* **8**:181-189.
- Paul, S.K. and Duara, R.K.** (1991). Combining ability studies in maize (*Zea mays* L.). *Intl. J. Tropic. Agric.*, **9**: 250-254.
- Pavan, R., Gangashetty, P. and Mallikarjuna, N.M.** (2011). General and specific combining ability studies in single cross hybrids of maize (*Zea mays* L.). *Current Biotica*. **5**(2): 196-208.
- Poneleit, C.G. and Bauman, L.F.** (1970). Diallel analysis of fatty acids in corn oil. *Crop Sci.* **10**: 338-341.
- Premlatha, M. and Kalamani, A.** (2010). Heterosis and combining ability studies for grain yield and growth characters in maize (*Zea mays* L.). *Indian Journal of Agri. Res.* **44**: 62- 65.
- Singh, P.K., Singh, N., Singh, A.K., Shahi, J.P. and Rao, M.** (2013). Heterosis in relation to combining ability in quality protein maize (*Zea mays* L.). *Biolife*. **1**(2): 65-69.
- Sproule, A. M., Saldivar, S. O., Bockholt, A. J., Rooney, L. W. and Knabe, D. A.** (1988). Nutritional evaluation of tortillas and tortilla chips from quality protein maize. *Cereal Foods World*. **33**(2):233-236.
- Vasal, S.K.** (1999). Quality Protein Maize story. In workshop on improving Human nutrition through Agriculture-The role of international agricultural research, October 5-7, IRRI, Philippines.