

## ISOLATION AND CHARACTERIZATION OF VARIOUS FUNGAL STRAINS AS PRIMARY COLONISER FROM WHEAT STRAW AT VARYING NITROGEN CONCENTRATIONS

Shahnaaz Khatoon<sup>1\*</sup> S.C. Jain<sup>1</sup> and M.U. Charaya<sup>2</sup>

<sup>1</sup>Department of Botany, D. N. (P.G.) College Meerut, Uttar Pradesh 250005

<sup>2</sup>Microbiology Laboratory, Department of Botany, Chaudhary Charan Singh University Meerut Uttar Pradesh 250005

Email: [Shahnaazkhatoon@gmail.com](mailto:Shahnaazkhatoon@gmail.com)

Received-08.10.2017, Revised-25.10.2017

**Abstract:** The present study was undertaken with an aim to search for the fungal strains, which have the potential to efficiently decompose wheat straw with high C:N ratios. Identification and characterization of these microbial species is important to study their decomposition potential for use in soil fertility management. A total 19 strains of fungal primary colonizers were isolated from a sample of wheat straw. Out of these, one belonged to Zygomycota while the remaining 18 belonged to Deuteromycota. *Alternaria*, *Aspergillus*, *Cladosporium*, *Helminthosporium*, *Stachybotrys*, *Fusarium* and *Penicillium* were the most frequently isolated genera at low nitrogen concentration. Isolated strains at low nitrogen concentration seem to be the most probable candidates, as initial primary bio inoculants, for hastening the decomposition of wheat straw. The results of this study suggest the possibility of utilizing fungal inoculants as an integrated component of microbe-based strategies for biotechnological management of wheat straw.

**Keywords:** Wheat Straw, Microorganisms, Isolation, Decomposition, Serial dilution, Fertilizers

### INTRODUCTION

Wheat is grown basically for the grains. But the utility of the remaining parts of the wheat plant cannot be ignored. A lot of wheat straw is produced in wheat-growing belts in the world. A significant proportion of wheat straw has been in use as feed for ruminants because of its abundance and low cost (Viola *et al.* 2008; Balset *et al.*, 2010; Manriquez *et al.* 2016). However, it is also used for the production of pulp and paper (Zhao *et al.* 2004), strawboards (Deswarteet *et al.*, 2007), textiles and composites (Avella *et al.*, 1995; Reddy *et al.* 2007), plastics (Avella *et al.*, 1995) and for the removal of metals in wastewater industry (Kumar *et al.*, 2000; Doan *et al.*, 2008) and hybrid composite materials (Yu *et al.* 2016).

Incidentally, a major portion of straw is burnt in the field itself (Gupta *et al.*, 2004). This results, on one hand, in a waste of organic sources in soil affecting C:N ratio and biota; and on other hand, leads to global warming and environmental problems (Badrinath *et al.* 2006; Sastre *et al.* 2015). Keeping in mind the harmful effects of burning straw in the field as well as the convenience of the farmer, economical, environment-friendly and low labour strategies should be adopted for effective utilization of the straw.

Primary colonizers are the microorganisms which play an important role in the initiation of decomposition. These organisms span parasitic as well as saprophytic phases. They possess cellulolytic (Sajith *et al.* 2015) and pectolytic activity at low nitrogen level, and are able to grow at faster rate on a comparatively drier resource (Charaya *et al.* 2005,

2006), (Chauhan *et al.*, 2006) and (Rani, 2008) also found that majority of the primary colonizers of plant litter possessed weak parasitic tendency.

Organic matter plays a unique role in soil fertility (Zhao *et al.* 2016). It acts as a sink as well as a source for nutrient. It prevents environmental pollution and the loss of nutrients. Above all, it helps in maintaining nutrients balance in the soil which is the basic attribute for sustainability (Raman, 2005). In a natural ecosystem, entire biomass of the plants is returned to the soil after the death of the plants through the process of decomposition. However, in agro-ecosystems, a significant proportion of the biomass is removed from the soil. Hence, intensive cultivation of crop requires massive application of synthetic fertilizers to compensate for the loss of nutrients from the soil as a result of their removal by the agricultural crops.

However, the prohibitive cost of chemical fertilizers as also numerous environmental problems associated with their production and use have prompted agricultural scientists to look for better alternatives. It is being gradually realized that organic wastes and biological sources of nutrients are better alternatives and these may serve as substitutes for inorganic fertilizers to a considerable extent, if not absolutely (Shukla and Mathur, 2000). Wheat straw provides one such alternative. The application of biodegraded products of straw into soil has enormous potential to recycle nutrients and maintain soil fertility (Gand *et al.*, 2006; Zhang *et al.* 2015, Rahman *et al.* 2016).

Straw contains approximately 0.5% nitrogen and 40% carbon. Straw, when subjected to colonization by fungi, has only 0.5 units of nitrogen to offer to the fungi which generally require 1.2 to 1.6 units for

\*Corresponding Author

growth; thus, a deficit of 0.7 to 1.1 units of nitrogen appears in the environment (Alexander, 1977). Many workers including Park (1976) pointed out that the fungi which show high degree of cellulolytic ability under laboratory conditions fail to colonize and degrade the plant residues in a correspondingly effective manner.

Hence, it is important to take into account the nitrogen level of resources to be degraded in the selection of effective decomposers. Since wheat straw has low nitrogen content, it should be preferable to use those organisms for the purpose of decomposition which can grow, reproduce and carry out polysaccharolytic activity at low nitrogen levels. In other words, those microorganisms which can operate the process of decomposition with lesser units of nitrogen would be more suitable as primary colonizers or primary bioinoculants.

However, van Fassen and van Dijk (1979) have demonstrated that hot composting of straw leads to considerable losses of nitrogen in the form of ammonia also, and therefore only relatively small gains in nitrogen appear to take place during decomposition. If it is so, the secondary colonizers also have to face the deficiency of nitrogen. The addition of nitrogen to crop residues has been reported to enhance the rate of decomposition of crop residues but only when a large amount of residues with low nitrogen was decomposing. Additional supply of nitrogen is believed to enhance the growth of decomposers (Fan *et al.*, 1981; Yadav, 1987; Mary *et al.* 1996; Hu *et al.* 2015; Wang *et al.* 2015 and Maaroufi *et al.* 2017).

Primary colonizers are the microorganisms which play an important role in the commencement of decomposition of crop residues. Identification and characterization of these microbial species is important for studying their decomposition potential vis a vis soil fertility management. In the present study an attempt has been made towards isolation and characterization of fungal strains which can survive and flourish at low nitrogen concentrations or higher C/N ratios.

## MATERIAL AND METHOD

### Isolation of Fungi

Freshly harvested wheat straw was collected from agricultural fields situated at village Khardoni, Meerut. The samples were collected aseptically in fresh polythene bags and brought to the laboratory for further studies. Serial dilution plate method (Waksman, 1927) was used to isolate the fungi from wheat straw sample. 1 g of the sample was placed in 250 ml of sterile water and stirred for fifteen minutes using a magnetic stirrer to get the stock solution. 10 ml of this solution were immediately transferred to a conical flask containing 90 ml of sterile distilled water to get a suspension of 1:10 dilution. This suspension was used for the preparation of further

serial dilutions (1:100, 1:1000). From the suspension of each dilution (1:10, 1:100, 1:1,000), 1 ml aliquots were transferred to each of a set of three Petri dishes followed by the addition of approximately 20 ml of cooled (45°C) and sterilized culture medium. Czapek'sDox Agar medium (Raper and Thom, 1949) with 30 ppm of rose bengal and 30 mg of streptomycin was used for this purpose. This medium served as control and was designated as N. The media with 2/3 (two third), 1/2 (half) and 1/3 (one third) concentrations (i.e. 1.5 g/l, 1.0 g/l and 0.67 g/l respectively) of nitrogen (sodium nitrate) as compared to control (2.0 g/l) were also prepared. The media with 2/3, 1/2 and 1/3 concentrations of nitrogen were designated as N/1.5, N/2 and N/3 respectively.

### Records of the Fungi Isolated

Isolation and identification of the fungi was done in the microbiology laboratory of the Department of Botany, CCS University, Meerut, India. The Petri dishes were observed from the third day itself when fast-growing fungi started appearing in the Petri dishes. The slow-growing fungi were transferred onto other Petri dishes just after their appearance to prevent them from being overrun by the fast growing fungi. A complete record of the fungal species and their numbers in cfus (cfus: colony forming units) in the Petri dishes was maintained. The identification of the fungal species was done on the basis of their morphology and cultural characteristics following Gilman (1957), Barnett and Hunter (1972), Subramanian (1971), Ellis (1971, 1976), Domsch and Gams (1972), Domsch *et al.* (1980), Nagmani *et al.* (2006), Venkateswarlu *et al.* (2015), Singh *et al.* (2015).

### Purification and Maintenance of Cultures

The fungal strains were purified by hyphal cut method and streak plate method. The pure cultures were maintained on PDA medium (Riker and Riker, 1936) on slants and were stored in a refrigerator.

## RESULT AND DISCUSSION

The current practice of using media having low C:N ratio for the isolation of microflora colonizing the resources with much higher C:N level in most biological laboratories is not appropriate as it might not present a true picture of the colonizer microorganisms. There are greater chances that the microbes present on the resource would be favored by the medium having higher nitrogen levels, thus distorting the picture completely by tilting the balance against the active microorganisms not capable of growing fast enough on the medium with high nitrogen and in favor of those microbes which are not actually active on the litter but grow better on the medium employed. Therefore, it would be worthwhile to use media with the C:N ratio comparable to that in the resource under study. The dominance of Deuteromycota observed in the present



<i>P. spinulosum</i>	—	—	—	1	11.11	I	—	—	—	—	—	—
<i>Rhizopus sp.</i>	3	22.22	II	—	—	—	1	11.11	I	—	—	—
<i>Stachybotrys atra</i>	5	33.33	II	4	33.33	II	3	22.22	II	1	11.11	I
<b>Total Isolates</b>	<b>33</b>			<b>26</b>			<b>35</b>			<b>23</b>		
<b>Total Species</b>	<b>11</b>			<b>6</b>			<b>8</b>			<b>11</b>		

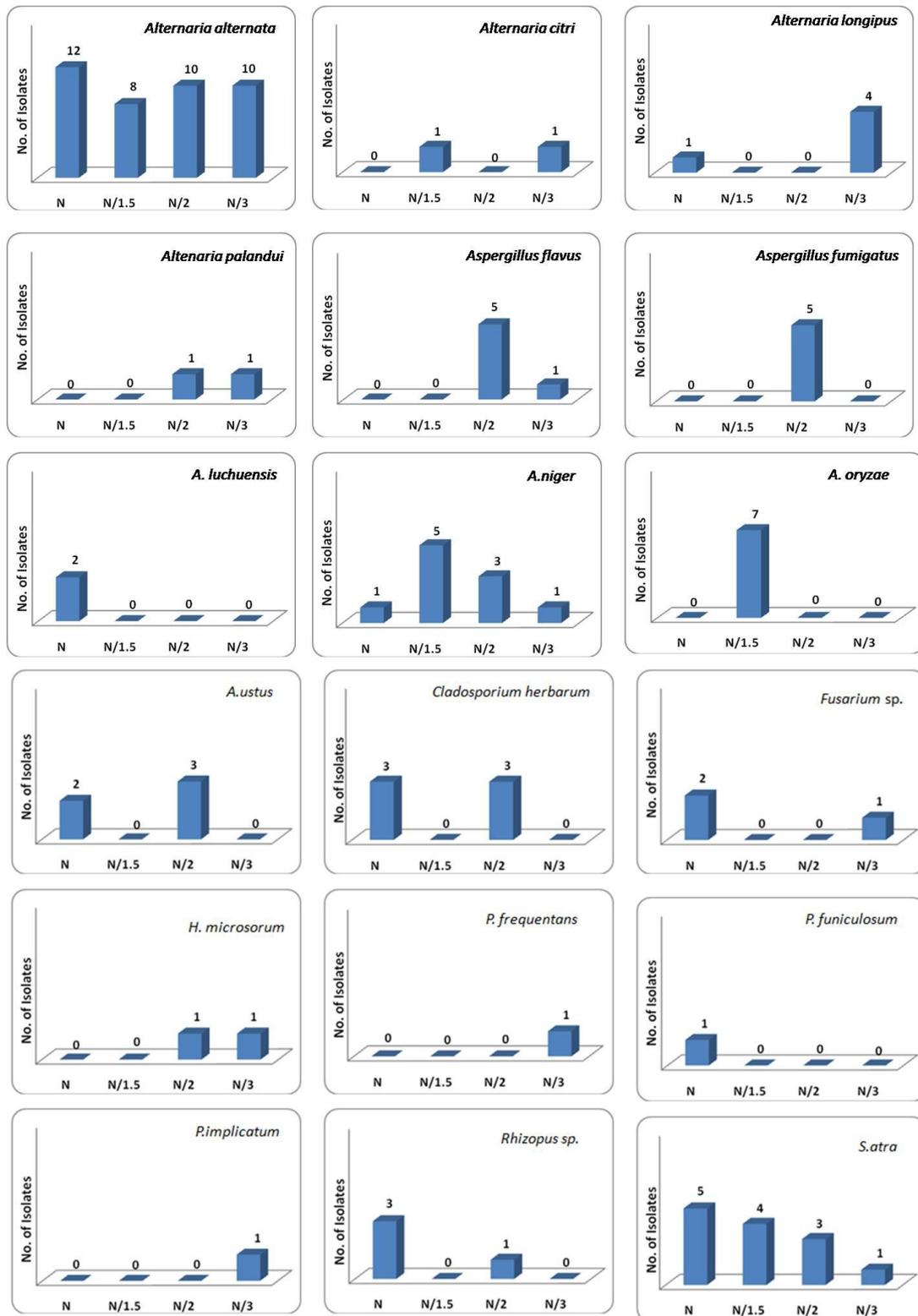


Fig 1: A comparison of the Total Isolate at different nitrogen levels

Total 35 isolates at N/2 media belonging to 8 fungal species were identified as *A. alternate* (10), *A. palandui* (1), *A. flavus* (5), *A. fumigates* (5), *A. niger* (3), *A. ustus* (3), *C. herbarum* (3) and *H. microsorum* (1). *A. citri*, *A. longipes*, *A. luchuensis*, *A. oryzae*, *Fusarium* sp. *P. frequentans*, *P. funiculosum*, *P. paxilli* and *P. spinulosum* were not obtained at this nitrogen concentration.

Total 23 fungal isolates belonging to 11 species were obtained at N/3 media and identified as *A. alternata* (10), *A. citri* (1), *A. longipes* (4), *A. palandui* (1), *Aspergillus flavus* (1), *A. niger* (1), *Fusarium* sp. (1), *H. microsorum* (1), *Penicillium frequentans* (1), *P. implicatum* (1) and *Stachybotrys atra* (1). *A. fumigates*, *A. luchuensis*, *A. oryzae*, *A. ustus*, *C. herbarum*, *P. funiculosum*, *P. paxilli*, *P. spinulosum* and *Rhizopus* sp. were not obtained at N/3 media. It was apparent from the results that *Alternaria alternata* was abundant at all nitrogen concentrations used in this study (table 1 and fig. 1).

## CONCLUSION

The present study describes isolation and identification of fungal colonizers growing actively at low nitrogen concentrations, having potential decomposing activity and their enzyme make-up. Attempt has also been made to identify the fungal strains which respond favourably to supplementation of the resource with additional nitrogen. The results suggest the possibility of utilizing fungal inoculants as an integrated component of microbe based strategies for biotechnological management of agricultural wastes. Czapek's media with different concentrations of nitrogen were prepared to the study the effect of different nitrogen levels on radial growth of selected fungal species Eighteen fungal species viz. *Alternaria alternata*, *A. citri*, *A. longipes*, *A. palandui*, *Aspergillus flavus*, *A. fumigatus*, *A. luchuensis*, *A. niger*, *A. oryzae*, *A. ustus*, *Cladosporium herbarum*, *Fusarium* sp., *Helminthosporium microsorum*, *Penicillium frequentans*, *P. funiculosum*, *P. implicatum*, *P. paxilli*, *P. spinulosum* and *Stachybotrys atra* were taken for studying effect of nitrogen and results of this study would be communicated in a subsequent paper.

## ACKNOWLEDGEMENT

Shahnaaz Khaton expresses her gratitude to Ph.D supervisors, Dr. M.U. Charaya and Dr. S.C. Jain, who have been a constant source of guidance and motivation throughout the course of this study.

## REFERENCES

**Alexander, M.** (1977). Introduction to Soil Microbiology. 2<sup>nd</sup> Ed. John Wiley and Sons,

Reprinted by Wiley Eastern Limited, New Delhi, Bangalore, Bombay, Calcutta.

**Avella, M., Bozzi, C., dell'Erba, R., Focher, B., Marzetti, A. and Martuscelli, E.** (1995). Steam exploded wheat straw fibers as reinforcing material for polypropylene-based composites. *Die Angew and teakro molekulare Chemie.* 233: 149–166.

**Badrinath, K.V.S., Kiranchand, T.R. and Prasad, Krishna** (2006). Agriculture crop burning in the Indo-Gangetic plains- a study using IRS-P6AWiFS satellite data. *Curr. Sci.* 91, 85–89.

**Bals, B., Murnen, H., Allen, M. and Dale, B.** (2010). Ammonia fiber expansion (AFEX) treatment of eleven different forages: Improvements to fiber digestibility in vitro. *Animal Feed Science Technology* 155: 147–155. Mary, B., Recous, Darwis, D., Robin, D., 1996. Interactions between decomposition of plant residues and nitrogen cycling in soil. *Plant and Soil.* 181, 71–82.

**Barnett, H.L. and Hunter, B.B.** (1972). Illustrated Genera of Imperfect Fungi. Burgess Publishing Company, Minneapolis, Minnesota.

**Charaya, M.U.** (2006). Successive microbial colonization of wheat straw decomposing under different situations. *Jour. Ind. Bot. Soc.* 85, 121–134.

**Charaya, M.U. and Mehrotra, R.S.** (2005). Fungal decomposition of plant litter: some attributes of primary colonizers. Proc. Symposium on challenging problem in Mycology and Plant pathology. May 2-3, 2005, Jammu (J &K); pp. 29–31.

**Charaya, M.U. and Mehrotra, R.S.** (2005). Fungal decomposition of plant litter: some attributes of primary colonizers. Proc. Symposium on challenging problem in Mycology and Plant pathology. May 2-3, 2005, Jammu (J &K); pp. 29–31.

**Chauhan, K.** (2006). Studies on potential of fungi to decompose sugarcane trash in relation to varying nitrogen levels. Ph.D. Thesis C.C.S. University, Meerut.

**Chauhan, K.** (2006). Studies on potential of fungi to decompose sugarcane trash in relation to varying nitrogen levels. Ph.D. Thesis C.C.S. University, Meerut.

**Deswarte, F.E.I., Clark, J.H., Wilson, A.J., Hardy, J.J.E., Marriott, R., Chahal, S.P., Jackson, C., Heslop, G., Birkett, M., Bruce, T.J. and Whiteley, G.** (2007). Toward an integrated straw-based biorefinery. *Biofuels, Bioproducts and Biorefining,* 1, 245–254.

**Dickinson, C.H. and Pugh, G.J.F.** (1974). *Biology of Plant Litter Decomposition.* Vols. I and II. Academic Press, London and New York.

**Doan, M., Ingrid, H., Franke-Whittle, I., Insam, H., Chen, Y. and Hadar, Y.** (2008). Molecular analysis of bacterial community succession during prolonged compost curing. *FEMS Microbiology Ecology* 65, 133–144.

**Domsch, K.H. and Gams, W.** (1972). Fungi in Agricultural Soils. Longman, London.

- Domsch, K.H., Gams, W. and Anderson, T.** (1980). *Compendium of Soil Fungi*. Vol. I & II. Academic Press, London, New York, Sydney.
- Dube, V.P., Charaya, M.U. and Modi, P.** (1980). Ecological and *in vitro* studies on the soil mycoflora of mango orchards. *Proc. Ind. Acad. Sci. (Plant Sci.)* 82, 157–160.
- Ellis, M.B.** (1971). *Dematiaceoushyphomycetes*. Commonwealth Mycological Institute, Kew, Surrey, England.
- Ellis, M.B.** (1976). *More Dematiaceoushyphomycetes*. Commonwealth Mycological Institute, Kew, Surrey, England.
- Fan, L.T., Lee, Y.H. and Beardmore, D.H.** (1981). The influence of major structural features of cellulose on rate of enzymatic hydrolysis. *Biotechnol. Bioeng.* 23: 419–442.
- Gaind, S., Pandey, A.K. and Lata** (2005). Biodegradation study of crop residues as affected by exogenous inorganic nitrogen and fungal inoculants. *J. Basic Microbiol.* 45, 301–311.
- Galloway, L.D.** (1935). Indian Soil fungi. *Ind. J. Agric. Sci.* 6, 578–585.
- Guleri, S., Saxena, S., Sharma, P., Malik, N. and Thapliyal, M.** (2016). Occurrence and Diversity of Soil Mycoflora in Some Selected Brassica Growing Agricultural Fields of Dehradun District of Uttarakhand Himalaya. *Int. J. Pure App. Biosci.* 4, 253-264.
- Gupta, P.K., Sahai, S., Singh, N., Dixit, C.K., Singh, D.P., Sharma, C., Tiwari, M.K., Gupta, R.K. and Garg, S.C.** (2004). Residue burning in rice wheat croppingsystem: causes and implications. *Curr Sci.* 87, 13–15.
- Hayes, W.A. and Lim, W.G.** (1979). Wheat and rice straw composts and mushroom production. In “Straw decay and effect of its disposal and utilization” (Ed. Gronbard, E.); John Willey and sons, Chichester, New York, Brisbane, Toronto pp. 83–93.
- Hudson, H.J.** (1968). The ecology of fungi on plant remains above the soil. *New Phytol.* 67, 837–874.
- Jensen, H.L.** (1931). The fungus flora of the soil. *Soil Sci.* 31, 123–158.
- Kumar, A., Rao, N.N. and Kaul, S.N.** (2000). Alkali-treated straw and insoluble straw xanthate as low cost adsorbents for heavy metal removal preparation, characterization and application. *Biores. Tech.* 71, 133–142.
- Li, H.C., Hu, Y.L., Mao, R., Zhao, Q. and Zeng, D.H.** (2015). Effects of Nitrogen Addition on Litter Decomposition and CO<sub>2</sub> Release: Considering Changes in Litter Quantity. *PLoS ONE* 10(12), e0144665.
- Maaroufi, N.I., Nordin, A., Palmqvist, K. and Gundale, M.J.** (2017). Nitrogen enrichment impacts on boreal litter decomposition are driven by changes in soil microbiota rather than litter quality. *Scientific Reports*, 7, 4083.
- Manríquez, O. M., Montano, M. F., Calderon, J. F., Valdez, J. A., Chirino, J. O., Gonzalez, V. M. and Zinn, R. A.** (2016). Influence of Wheat Straw Pelletizing and Inclusion Rate in Dry Rolled or Steam-flaked Corn-based Finishing Diets on Characteristics of Digestion for Feedlot Cattle. *Asian-Australas J. Anim Sci.* 29, 823–829.
- Nagamani, A., Kunwar, K.I.O. and Manoharachary, C.** (2006). *Handbook of soil fungi*. I.K. International Pvt. Ltd. New Delhi.
- Park, D.** (1976). Carbon and nitrogen levels as factors influencing fungal decomposers. In: *The Role of Terrestrial and Aquatic organisms in Decomposition process* (Eds. Anderson, J.M. and A. Macfadyen). Blackwell Scientific Publications. Oxford, London Edinburgh, Melbourne pp. 41–59.
- Rahman, F., Rahman, M., Rahman, G.K.M., Saleque, M.A., Sakhawat Hossain A.T.M. and Md Giashuddin Miah.** (2016). Effect of organic and inorganic fertilizers and rice straw on carbon sequestration and soil fertility under a rice–rice cropping pattern. *Carbon Management*, 7, 41-53.
- Raman, K.V.** (2005). Sustaining soil fertility. *The Hindu Survey of Indian Agriculture pp.* 165–167.
- Rani, P.** (2008). Studies on the fungal decomposition of sugarcane trash. Ph.D. Thesis C.C.S. University, Meerut.
- Raper, K. B. and Thom** (1949). *A Manual of Penicillia*. Williams Wilkins Co., Baltimore, Md.
- Reddy N. and Yang, Y.** (2007). Preparation and characterization of long natural cellulose fibers from wheat straw. *J. Agric. Food Chem.* 55, 8570–8575.
- Riker, A. J. and Riker, R.S.** (1936). *Introduction to research on plant diseases*. John S. Swift Co., St. Louis, Mo.
- Runzhou Huang, Min Yu., Chunxia He, Qinglin Wu, and Xueni Zhao** (2016). Hybrid Composites from Wheat Straw, Inorganic Filler, and Recycled Polypropylene: Morphology and Mechanical and Thermal Expansion Performance. *Int. J. Polym. Sci.* 12 pages.
- Sajith, S., Priji, P., Sreedevi, S. and Benjamin, S.** (2016). An Overview on Fungal Cellulases with an Industrial Perspective. *J Nutr Food Sci* 6:461.
- Sastre, C.M., González-Arechavala, Y. and Santos, A.M.** (2015). Global warming and energy yield evaluation of Spanish wheat straw electricity generation - A LCA that takes into account parameter uncertainty and variability. *Appl. Energy.* 154, 900–911.
- Sen, S. and Charaya, M.U.** (2010). Copper-tolerant microfungi for bioremediation. *Prog. Agric.* 10, 66–71.
- Sen, S., Charaya, M.U. and Singh, P.B.** (2009). Screening of soil for lead tolerant fungi. *Indian J. Plant Genet. Resour.* 22, 191–194.
- Shukla, L. and Mathur, R.S.** (2000). Effect of biodegradation sugarcane trash on yield of nutrient uptake by wheat crop. *Journal of the Indian Society of Soil Science*, 48, 520–522.

- Singh, P.N. and Charaya, M.U.** (1975). Soil fungi of sugarcane field at Meerut. Distribution of soil mycoflora. *Geobios.* 2, 40–43.
- Singh, R.** (2004). Studies on the fungal decomposition of above ground residues of wheat crop. Ph.D. Thesis, C.C.S. University, India.
- Singh, R., Charaya, M.U., Shukla, L., G. Shukla., Kumar, A. and Rani, A.** (2015). Lignocellulolytic Potentials of *Aspergillus terreus* for Management of Wheat Crop Residues. *J. Acad and Ind. Res.* 3, 453-455.
- Subramanian, C.V.** (1971). *Hyphomycetes*. Indian Council of Agricultural Research, New Delhi.
- Tiwari, A.** (2010). Studies on the mitigation of metal pollution through bioadsorption by soilfungi. Ph.D. Thesis, Department of Botany, C.C.S. University, Meerut.
- Tiwari, A. and Charaya, M.U.** (2006). Effect of chromium sulphate on soil mycobiota. *Bio-Sci. Res. Bull.* 22, 53–56.
- Van Fassen, H.G. and Van Dijk, H.** (1979). Nitrogen conversions during the composting of manure/straw mixtures. In: straw decay and its effect on disposal and utilization (Ed. Grossbard, E.); John Willey and Sons, Chichester, N.Y., Brisbane, Toronto, pp. 113–120.
- Venkateswarlu, N., Sireesha, O., Aishwayra, S., Vijaya, T. and Sriramulu. A.** (2015). Isolation, screening of rhizosphere fungi antagonistic to rice stem rot disease pathogen sclerotium oryzae catt. *Asian J Pharm Clin Res*, 8, 54-57.
- Viola, E., Zimbardi, F., Cardinale, M., Cardinale, G., Braccio, G. and Gambacorta, E.** (2008). Processing cereal straws by steam explosion in a pilot plant to enhance digestability in ruminants. *Biores. Technol.* 99, 681–689.
- Waksman, S.A. and Tenney, F.G.** (1927). The composition of natural organic materials and their decomposition in the soil. I methods of quantitative analysis of plant materials. *Soil Sci.* 24, 275–283.
- Wang, J., Wensheng, Bu., Zhao, Bo., Zhao, X., Zhang, C., Juan, Fan. and Gadov, K.V.** (2015). Effects of Nitrogen Addition on Leaf Decomposition of Single-Species and Litter Mixture in *Pinus tabulaeformis* Forest. *Forests* 6, 4462–4476.
- Wei, T., Zhang, P., Wang, K., Ding, R., Yang, B. and Nie, J.** (2015). Effects of Wheat Straw Incorporation on the Availability of Soil Nutrients and Enzyme Activities in Semiarid Areas. *PLoS ONE* 10(4), e0120994.
- Yadav, J.S.** (1987). Influence of nutritional supplementation on solid substrate termination of wheat straw with an alkalophilic white rot fungus (*Coprinus* spp.). *App. Microbiol. Technol.* 26, 474–478.
- Zhao J., Li X., Qu Y. and Gao, P.** (2004). Alkaline peroxide mechanical pulping of wheat straw with enzyme treatment. *Appl. Biochem. Biotechnol.* 112, 12–23. Raper, K. B. and Thom (1949). *A Manual of Penicillia*. Williams Wilkins Co., Baltimore, Md.
- Zhao, Ya-Nan., Xin-Hua, He., Xing-Cheng, Huang., Yue-Qiang, Zhang. and Xiao-Jun, Shi.** (2016). Increasing Soil Organic Matter Enhances Inherent Soil Productivity while Offsetting Fertilization Effect under a Rice Cropping System. *Sustainability*, 8, 879.

