EFFECT OF ACCELERATED AGEING VARIABLES ON VARIOUS SEEDS QUALITY PARAMETER IN BARLEY (*HORDEUM VULGARE* L.)

D.S. Chauhan* and D.P. Deswal

Department of Seed Science and Technology CCS Haryana Agricultural University, Hisar Email: davender_<u>chauhan@rediffmail.com</u>

Received-08.09.2019, Revised-26.09.2019

Abstract: Accelerated Ageing test has good Correlation with field emergence and storage potential of Seeds. In This regard study was conducted to standardized accelerated ageing duration along with temperature variables by using six barley genotypes Viz.ALFA-93, BH-393, BH-75, BCU-3, RD-2552 and K-551. All Six varieties was divided in two seed lots subjected to accelerated ageing at two time variables $42\pm1^{\circ}$ C and $45\pm1^{\circ}$ C for 48 and 72 hrs separately on 100% relative humidity. Seeds were exposed to accelerated ageing conditions at two temperature 42°C and 45°C for different time intervals 48, 72 hrs; the viability reduced significantly ranging from 40 to 60%. In the present study genotypes BH-393, BCU-73 were observed to be tolerant to stress conditions during accelerated ageing at 42°C for 48 hrs. whereas Genotypes BH-393 and RD-2552 were observed most tolerant to stress conditions during accelerated ageing at 45°C for 48 hrs in present study, the genotypes showing high resistance to accelerated ageing test were also showing high standard germination in fresh seed ranging from 94 to 100%. Results revealed that the germination percentage for fresh seed lot was above the minimum seed certification standard whereas in accelerated aged seed lots, the standard germination, vigor index, rate of germination and dehydrogenase activity declined significantly for all the six genotypes. All the genotypes were found significantly different for vigor and viability tests. After accelerated ageing, the genotypes, BH-393 and BCU-73 were having significantly higher standard germination, vigor, rate of germination and enzymatic activity as compared to other genotypes indicating their superiority over others. A higher electrical conductivity of accelerated aged seed in barley further confirmed the accelerated ageing results in to increase permeability of cell membrane and release of food reserves.

Keywords: Accelerated ageing, Standard Germination, Barley seed

REFERENCES

Abdul-Baki, A.A. (1980). Biochemical aspects of seed vigor. Hort Science, 15: 765-771.

Abdul-Baki, A.A. and Anderson, J.D. (1972). Physiological and biochemical deterioration of seeds. In: Koziowski, T.T. (ed.) *Seed Biology*, Vol. 2, 283-315. Academic Press, New York.

Anderson, J.B. and Abdul-Baki, A.A. (1973). Vigor determination in soyabean seeds by Multiply criteria. *Crop Sci.* 13: 630-633.

Andrade, R. V., Auzza, S.A.Z., Andreoli, C., Netto, D.A.M. and Oliveira, A.C. (2001). Seed quality evaluation of the maize single cross hybrid MS 200. *Ciencio-e- Agrotechnologio* 25(3): 576-582.

Bailly C. (2004). Active oxygen species and antioxidants in seed biology. Seed Sci Res, 14:93-107.

Copeland, L.O. and McDonald M.B. (2001). Principles of seed science and technology. 4th edition. Kluwer academic publishers, pp. 176.

Bailey, C., Banamar, A., Corbineau, F. and Come, D. (1996). Changes in mal on dealdehyde content and insuperoxide dimutase, catalase and glutothione reductase activities in sunflower seeds as related to deterioration during accelerated ageing. *Physiol. Plant.* 97: 104-110.

Basra, S.M.A., Ahmad, N., Khan, M.M., Iqbal, N. and Cheema, M.A. (2003). Assessment of cotton seed deterioration during Accelerated Ageing. *Seed Sci. and Technol.* 31(3): 531-540.

*Corresponding Author

Ching, T.M. (1973). Bio-chemical aspects of seed vigour. *Seed Sci. & Technol.*, 1:73-88.

Delouche, J.C. and Baskin, C.C. (1978). Accelerated ageing techniques for predicting the relative storability of seed lots. *Seed Sci. & Technol.* 15 : 427-452.

Freitas, R.A., Dias, D.C.P. S., Cecon, P. R., Reis, M.S. and Dias, L.A.S. (2002). Storability of cotton seeds predicted by vigour tests. *Seed Science and Tech.* 30(2): 402-410.

Gupta, P.K. and Agrawal, R.L. (1980). Determination of variety purity of paddy varieties by laboratory evaluation. *Oryza.* 25:310-314.

Goel, A., Goel, A.K. and Sheoran, I.S. (2002). Changes in Oxidative stress enzymes during artificial ageing in cotton (*Gossypium hirsutum L.*) seeds. J Plant Physiol, 160:1093-1100.

Ghasemi-Golezani, K., Salehian, H., Rahimzade-Khoee, F. and Moghadam, M. (1996). The effect of seed vigor on seedling emergence and yield of wheat. Natural Resources and Agricultural Sciences, 3: 58-48.

Ghazi, N., Karaki, A., Al-Ajam, A. and Othman, Y. (2007). Seed germination and early root growth of root growth of three barley cultivars as affected by temperature and water stress. American-Eurasian J Agri and Environ Sci, 2(2): 112-117.

Harrington, J.F. (1960). Thumb rules of drying seeds. *Crops and soils*.13: 16-17.

Indira, K., Gunnasekaram, M. and Prostath, D. (2000). Accelerated ageing test to predict the

storability of fenugreek seeds. Orissa I. Hort. 28(1): 34-37.

ISTA (1999). International rules for seed testing. *Seed Sci. and Technol.* 23 (Suppl.) 1-334.

Janmohammadi, M., Fallahnezhad, F., Golsha, M. and Mohammadi, H. (2008). Controlled ageing for storability assessment and predicting seedling early growth of canola cultivars (*Brassica napus* L.). ARPN J Agric Biol Sci, 3:22-26.

Kittock, D.L. and Law, A.G. (1968). Relationship of seedling vigour to respiration and tetazolium

chloride reduction by germinating wheat seeds. *Agron. J.* 1: 417-425.

McDonald, M.B. (2004). Orthodox seed deterioration and its repair, pp. 273-304.*In*: Handbook of Seed Physiology: Application to Agriculture,Benech-Arnold,R.L. and R.A. Sanchez (Eds.) Food product Press,Newyork.

McDonald, M.B. (1999). Seed deterioration: physiology, repair and assessment. Seed Sci Technol, 27: 177-237.

Moradi, A. and Younesi, O. (2009). Effects of Osmo- and Hydro-priming on Seed Parameters of Grain Sorghum (*Sorghum bicolor* L.). Australian Journal of Basic and Applied Sciences, 3(3): 1696-1700.

Murata, M., Roos, E.E. and Tsuchiya (1979). Relationship between loss of germinability and occurence of chromosomal abberations in artificial aged seeds of barley. *Barley Genetic Newsletter*. 9: 65-67.

Narwal, A.L. (1995). Studies on seed viability in okra (*Abelmoschus esculentus*) (L.) Moench). Ph.D. Thesis, CCS HAU, Hisar.

Pandey, P.K., Goyal, R.D., Parakash, V., Katiyar, R.P. and Singh, C.B. (1990). Association between laboratory vigor tests and field emergence in cucurbits. Seed Sci Res, 18: 40-43.

Saha, R.R. and Sultana, W. (2008). Influence of seed ageing on growth and yield of soybean. Bangla J Bot., 37:21-26.

Siadat, S.A., Moosavi, A. and Sharafizadeh, M. (2012). Effect of seed priming on antioxidant activity and germination characteristics of maize seeds under different aging treatments. Research Journal of Seed Science, 5(2): 51- 62.

Soltani, E., Kamkar, B., Galeshi, S. and Akram Ghaderi, F. (2008). The effect of seed deterioration on seed reserves depletion and heterotrophic seedling growth of wheat. Journal of Agricultural Sciences and Natural Resources, 15(1):13-17 (In Persian).

Singh, B., Singh, C.B. and Gupta, P.C. (2003). Influence of Seed Ageing in Vigna species. *Farm Science Journal*. 12(1): 4-7.

Steiner, J.J., Grabe, D.F. and Tulo, M. (1989). Single and multiple vigour tests for predicting seedling emergence of wheat. *Crop Sci.* 29: 782-786. Walters, C. (1998). Understanding the mechanisms and kinetics of seed aging. Seed Sci Res, 8:223-244.